



## Microbiology of the food chain: Horizontal method for the detection, enumeration and serotyping of *Salmonella* – Detection of *Salmonella* spp. AS 5013.10-2022

This standard is an adoption with national modification of ISO 6579-1:2017. Microbiology of the food chain – Horizontal method for the detection, enumeration and serotyping of *Salmonella* – Part 1: Detection of *Salmonella* spp. and its Amendment No. 1. This standard replaces AS 5013.10.2009.

### SCOPE

This method is applicable to:

- products intended for human consumption (including raw meats and carcass swab and rinse samples) and the feeding of animals.
- environmental samples in the area of food production and food handling.

### PRINCIPLES

*Salmonella* Hofit is used as the positive control for this method. The detection of *Salmonella* spp. is broken down into four stages:

- **Pre-enrichment in non-selective liquid medium**  
For meat and meat products a 1:10 dilution of the sample is enriched in buffered peptone water at  $36 \pm 2^\circ\text{C}$  for  $18 \text{ h} \pm 2 \text{ h}$ . Buffered peptone water should be warmed to room temperature or to  $36 \pm 2^\circ\text{C}$  for large volumes (i.e. >225 mL). For carcass sponges, buffered peptone water is added to the moistened sponge to bring the total volume to 60-100 ml and the sample incubated at  $36 \pm 2^\circ\text{C}$  for  $18 \text{ h} \pm 2 \text{ h}$ . In the case of sponges BPW need not be warmed to room temperature before being used to re-hydrate the sponge, for all subsequent additions BPW should be warmed to room temperature.
- **Enrichment in selective liquid medium**  
Culture from the pre-enrichment broth is inoculated into Rappaport-Vassiliadis medium with soya (RVS broth) and Muller-Kauffmann tetrathionate/novobiocin broth (MKTTn broth, pH 8.0  $\pm$  0.2 at  $25^\circ\text{C}$ ). The RVS broth is incubated at  $41.5 \pm 1^\circ\text{C}$  for  $24 \text{ h} \pm 3 \text{ h}$  and the MKTTn broth at  $37 \pm 1^\circ\text{C}$  for  $24 \text{ h} \pm 3 \text{ h}$ .
- **Plating out and identification**  
Cultures obtained from the selective enrichment are streaked onto two selective media:
  - Xylose lysine deoxycholate agar (XLD agar)
  - And, for testing as part of export certification, any other solid selective medium that is complementary to XLD and able to detect  $\text{H}_2\text{S}$  negative serovars of *Salmonella* e.g. Brilliant Green Agar (BGA).XLD agar is incubated at  $37 \pm 1^\circ\text{C}$  and examined after  $24 \text{ h} \pm 3 \text{ h}$ . The second agar is incubated according to the manufacturer's recommendations. The department does not require the use of duplicate 90 to 100 mm Petri dishes or a single 140 mm Petri dishes. Single 90 to 100 mm Petri dishes can be used. Confirmation can be directly off the selective agar if well isolated colonies are available.
- **Confirmation of *Salmonella***  
Colonies (maximum of 20) of presumptive *Salmonella* (subcultured on to nutrient agar if necessary) are confirmed by appropriate biochemical tests, as detailed in AS 5013.10 (2022). Preliminary confirmation at the isolating laboratory should include polyvalent O and H antisera. Rapid biochemical identification kits described in AOAC 978.24, AOAC 989.12, AOAC 991.13 and AOAC 2017.09 can be used for biochemical confirmation (section 9.5.3 of AS 5013.10). *Salmonella* isolates must be sent to a reference laboratory for serotyping.

**CHECKLIST**

<b>Pre-enrichment</b>	Is the buffered peptone water warmed to room temperature (to $36 \pm 2^\circ\text{C}$ for large quantities)?	_____
	Is the correct amount of enrichment broth used for the weight of sample analysed?	_____
	Is primary enrichment at $37 \pm 1^\circ\text{C}$ for 16-20h?	_____
	Is a positive control run with each batch of samples analysed?	_____
	Are reference cultures inoculated into primary enrichment broth at a level of 10 to 100 cells?	_____
<b>Selective-enrichment</b>	Is RVS broth sterilised at $115^\circ\text{C}$ for 15 minutes?	_____
	Is MKTTn broth boiled not autoclaved?	_____
	Is RVS incubated at $41.5 \pm 1^\circ\text{C}$ for $24 \pm 3$ h?	_____
	Is MKTTN incubated at $37 \pm 1^\circ\text{C}$ for $24 \pm 3$ h?	_____
	Are all complete selective liquid media prepared on the day of use (or is a validated shelf-life provided by the manufacturer)?	_____
<b>Selective plating</b>	What agars are used for isolation of suspect colonies?	_____
	Is the isolation of $\text{H}_2\text{S}$ negative strains considered in the laboratories methods manual and procedures?	_____
<b>Confirmation</b>	How are cultures obtained for biochemical tests (if not streaked onto Nutrient agar is a purity check carried out)?	_____
	Are approved rapid bio-chemical test kits used?	_____
	Does preliminary confirmation at the isolating laboratory include polyvalent O and H antisera?	_____
	Are biochemical tests used sufficient to identify <i>Salmonella</i> spp.?	_____
	Are all suspect <i>Salmonella</i> sent to a reference Laboratory to be serotyped?	_____